

# Linking common non-coding RNAs of human lung cancer and *M. tuberculosis*

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## Abstract:

Lung cancer and pulmonary tuberculosis caused by *Mycobacterium* are two major causes of deaths worldwide. Tuberculosis linked lung cancer is known. However, the precise molecular mechanism of *Mycobacterium* associated increased risk of lung cancer is not understood. We report 45 common human miRNAs deregulated in both pulmonary tuberculosis and lung cancer. We show that sRNA\_1096 and sRNA\_1414 from *M. tuberculosis* have sequence homology with human mir-21. Hence, the potential role of these three small non-coding RNAs in rifampicin resistance in pulmonary tuberculosis is implied. Further, the linking of sRNA\_1096 and sRNA\_1414 from *M. tuberculosis* with the host lung tumorigenesis is inferred. Nonetheless, further analysis and validation is required to associate these three non-coding RNAs with *Mycobacterium* associated increased risk of lung cancer.

**Keywords:** Genetic predisposition; lung cancer risk; microRNA; *Mycobacterium tuberculosis*, sRNA

## Background:

Viral involvements and their causal roles in oncology are well accepted for various cancers including ovarian neoplasms [1] hepatocellular carcinoma [2] and lung cancer [3] among others. Although, bacterial infections are not considered as major threats to cancer, yet a number of bacterial pathogens are reported to be associated with several cancers. Some examples include: *Mycoplasma* in prostate malignancy [4], *Robinsoniella* in pancreatic cancer [5], *S. typhi*, *H. bilis*, *H. hepaticus*, and *E. coli* in carcinoma of the gallbladder [6], *Chlamydia* in cervical cancer [7] and *Mycobacterium* in lung cancer [8-14].

Lung cancer is the leading cause of all cancer related deaths with a recently estimated 1.6 million deaths worldwide [15]. Similar to lung cancer, pulmonary tuberculosis caused by *M. tuberculosis* is a global health problem. It is one of the major causes of death amongst infectious diseases and according to WHO 2013 report, it is estimated that 9 million people are infected and 1.5 million died from tuberculosis in 2012 [16]. Several reports have

documented the co-existence of tuberculosis and lung cancer [14, 17-20], and pulmonary tuberculosis is a risk factor for developing lung cancer [17-20]. However, it is not yet fully established at the molecular level, how the *Mycobacterium* increases susceptibility to lung cancer. Some reports say that *M. tuberculosis* induces ROS mediated DNA damage pathway and produces epiregulin growth factor to induce cell proliferation [21]; while an other study indicates mechanisms along with COX-2 mediated activation of inflammatory pathway in *M. tuberculosis* associated carcinogenesis [22].

Bacterial small regulatory RNAs (sRNAs) are a class of small non-coding RNAs of 40-500 nt in length that regulate various essential patho-physiologies in bacteria such as outer membrane protein biogenesis, virulence, quorum sensing etc. sRNA functions through complementary base-pairing with 3'- or 5'-UTRs of target mRNAs to inhibit translation, alters activity of a protein by directly binding, and by mimicking RNA and DNA structures [23-25]. Several sRNAs have been identified or

predicted from *M. tuberculosis* [26–28] having probable role in pulmonary tuberculosis pathogenesis [26, 29, 30]. However, no report so far is available on sRNAs from *M. tuberculosis* having role in lung cancer. On the other hand, human micro RNAs (miRNAs) are small non-coding RNAs of 20–25 nt length that inhibit post-transcriptional gene regulation by complementary base pairing at the 3' -UTRs of target mRNAs and regulate various patho-physiological conditions including cell cycle regulation, cell differentiation, development, metabolism, aging, different types of cancers, metabolic disorders, and neuronal diseases etc. [31,32]. Several miRNAs have been implemented to be associated with lung cancer having causative roles and diagnostic potentials [33]. Similarly, a number of miRNAs are found deregulated in pulmonary tuberculosis patients [34–36].

Since the sRNAs and miRNAs are similar in structure and their mode of actions, and the miRNAs are associated with both pulmonary tuberculosis and lung cancer; we hypothesized that *Mycobacterium* sRNAs may be associated with lung cancer tumorigenesis. Since *Mycobacterium* infection is a risk factor in developing lung cancer, here we postulated a “Genetic remittance” hypothesis which presumes that *M. tuberculosis* sRNAs having similarity with human miRNAs (that are associated with either pulmonary tuberculosis or lung cancer or both diseases) are transmitted to the host during *Mycobacterium* infection, remained within the human, and act as a predisposition factor to increase the risk of lung cancer.

In this study, using *in silico* strategies, we aimed to understand (i) the role of *M. tuberculosis* sRNAs in lung carcinogenesis; (ii) if there are structurally and functionally similar *M. tuberculosis* sRNAs to human miRNAs responsible for pathogenesis in both pulmonary tuberculosis and lung cancer and their probable functions in these diseases; and (iii) proof of concept of our “Genetic remittance” hypothesis.

#### Methodology:

##### Collection of human miRNAs and *M. tuberculosis* siRNAs:

We used PubMed (<http://www.ncbi.nlm.nih.gov/pubmed>), miRegulome [32], miR2Disease [37], TUMIR [38], sRNAdb [39], and BSRD [40] databases to get the data. The data deposited in these databases during January 2006 to November 2015 were searched. In the first step, we collected all the validated deregulated miRNAs associated with lung cancer and pulmonary tuberculosis from published literature indexed in PubMed (<http://www.ncbi.nlm.nih.gov/pubmed>) and from databases such as miRegulome [32], miR2Disease [37], and TUMIR [38]. The common miRNAs associated with both the diseases were manually identified and listed out separately. Similar to miRNA, we collected all reported and novel *M. tuberculosis* sRNAs by means of PubMed literature mining and searching sRNA databases such as sRNAdb [39] and BSRD [40]. For PubMed search, specific key words such as “miRNA + *M. tuberculosis*”, “miRNA + tuberculosis”, “miRNA + lung cancer”, sRNA + *M. tuberculosis*”, “sRNA + tuberculosis” etc. were used.

##### Prediction of miRNAs that may function as sRNA or vice versa:

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To achieve this, we used a simple strategy. Since the sRNAs and miRNAs are very short in sequence, we presumed that small sequence motifs in these non-coding RNAs are very important for their specific functions. Therefore, we used comparative BLASTn (miRNA against sRNA) with default parameters in order to identify if there is any human miRNA (from the group of miRNAs common to both lung cancer and pulmonary tuberculosis) having sequence similarity to any *M. tuberculosis* sRNA, so they may have similar functions. From the BLASTn results, the specific motif sequences that are common in sRNAs and miRNAs were considered.

##### Functional annotation of *M. tuberculosis* sRNAs:

The functional annotation of the sRNAs was carried out using target based reverse annotation approaches following a modified protocol as described by Barh et al., 2013 [33]. In brief, we identified the validated targets of sRNA using sRNATarBase [41] database and RNApredator [42] was used to predict putative targets in *M. tuberculosis*, H37Rv. Top 100 targets were used for functional annotation by using the DAVID functional annotation tool [43]. Further, we presumed that, if there is a coding gene that has identity with sRNA; the function of the sRNA could be similar to that gene. Therefore, we performed sRNA BLASTn against *M. tuberculosis* genome using NCBI BLASTn server (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>) to identify if there is any coding sequence present in *M. tuberculosis* genome having sequence identity similar to that of sRNA. To check if the sRNA is targeting an essential gene of *M. tuberculosis*, H37Rv, we used Database of Essential Genes (DEG) [44] BLASTn and to check if the target could be a drug target, we used the strategy as described in [45]. Further, we did miRNA (that is having sequence similarity with sRNA) BLASTn against *M. tuberculosis* genome in NCBI BLASTn server with default parameters to check if any *M. tuberculosis* coding sequence is matching with the miRNA sequence. Since, identified miRNA in this way share sRNA sequences if there is a coding sequence that matches with the miRNA, we consider that, the matched coding sequence of the sRNA and the miRNA may have similar function.

##### Genetic remittance:

We performed BLASTn of the *M. tuberculosis* sRNAs having sequence similarity with human miRNA (that are associated with both pulmonary tuberculosis and lung cancer) against the human genome on NCBI BLASTn server in order to identify if there is any human coding sequence having identity with the sRNAs. Thus, common miRNAs that are deregulated both in pulmonary tuberculosis and lung cancer is observed. These observation indicate genetic remittance in tuberculosis associated lung cancer.

##### Results:

##### Common miRNAs in pulmonary tuberculosis and lung cancer:

From various literature and databases, we collected differential expression of 186 human miRNAs in pulmonary tuberculosis and 242 miRNAs in lung cancer patients. However, while we checked for the common miRNAs that are deregulated both in pulmonary tuberculosis and lung cancer, we found the number is only 45 (Supplementary Table S1, see supplementary data).

***M. tuberculosis* sRNA\_1096 and sRNA\_1414 shares hsa-mir-21 sequence:**

From the sRNADB [39] and BSRD [40] databases and literature [27, 46] we collected 120 reported *M. tuberculosis* sRNAs and their sequences. The comparative BLASTn between the 120 sRNAs and the common 45 human miRNAs in pulmonary tuberculosis and lung cancer revealed that the mature human miRNA hsa-mir-21-5p has very short sequence similarity with *M. tuberculosis* sRNA\_1096 and sRNA\_1414. Both these sRNAs are experimentally validated in *M. tuberculosis* [27]. Human hsa-mir-21-5p and *M. tuberculosis* sRNA\_1096 share a common sequence of GTTG/ GUUG, while the common sequence between hsa-mir-21-5p and *M. tuberculosis* sRNA\_1414 is ATCAG/ AUCAG (Supplementary Table S2, see supplementary data).

**Functional annotation of sRNA\_1096 and sRNA\_1414:**

To understand the functions of sRNA\_1096 and sRNA\_1414 in *M. tuberculosis*, first we used sRNATarBase [41] to search validated targets of these sRNAs. However we did not get any target from this database. Therefore, we used RNApredator [42] to predict the targets of these sRNAs. The top 100 targets based on the Z-score of RNApredator were further used for functional annotation using DAVID.

For sRNA\_1096, phosphate-binding protein pstS 2 (Z-score: -11.29) was found to be the best target. pstS 2 is an inorganic phosphate transmembrane transporter that is involved in two-component system and ABC transporters pathways. Further, among the top 100 targets, fourteen targets are PE PGRS family proteins (Supplementary Table S3, available with author). DAVID functional annotation analysis of the top 100 targets of sRNA\_1096 shows that most targets are membrane located and the two-component system is the top annotation cluster.

The top target of sRNA\_1414 is found to be glycyl-tRNA synthetase / glycine--tRNA ligase (glyS) (Z-score: -11.41) by RNApredator [42] (Supplementary Table S3, available with author). The drug transporter activity is ranked as the first annotation cluster and most targets are transmembrane proteins as observed through DAVID functional annotation analysis. According to the Database of Essential Genes (DEG) [44], the *M. tuberculosis* glyS is an essential gene in the pathogen but has 31% identity at protein level with human glycine--tRNA ligase as per NCBI human BLASTp.

When we performed sRNA\_1096 BLASTn against *M. tuberculosis* genome, a very short similarity "CCGTCACCGTTG" was observed with *M. tuberculosis* Arabinosyltransferase EmbC (embC) gene. The BLASTn of sRNA\_1414 with *M. tuberculosis* genome did not show any hit with any specific protein-coding gene of the pathogen.

**Common function of sRNA\_1096, sRNA\_1414, and hsa-mir-21:**

As in previous analysis we found there are sequence similarities among hsa-mir-21 and *M. tuberculosis* sRNA\_1096 and

sRNA\_1414; we performed a NCBI BLASTn of hsa-mir-21-5p against the *M. tuberculosis* genome. We observed that the *M. tuberculosis* rpoB gene that provides rifampin/ rifampicin resistance in *M. tuberculosis* has sequence similarity with mir-21 and has both the GTTG and ATCAG short-stretch sequences that are present in *M. tuberculosis* sRNA\_1096 and sRNA\_1414, respectively. Therefore, all these non-coding RNAs may be involved in rifampicin resistance in pulmonary tuberculosis.

**Support for "Genetic remittance" hypothesis:**

To support our "Genetic remittance" hypothesis, we tried to identify if there is any sequence match of sRNA\_1096 and sRNA\_1414 in human coding sequence. The BLASTn of sRNA\_1096 against human genome shows a hit with SH3GL1 (SH3-domain GRB2-like 1) and some sequence of sRNA\_1414 matches with human EPS8L1 (EPS8-like 1) and SORBS1 (Sorbin and SH3 domain containing 1). All these human genes are associated with tumorigenesis, thus providing a preliminary support to our hypothesis.

**Discussion:**

In this study we found that there could be correlations between lung cancer and pulmonary tuberculosis at the non-coding RNA level. *M. tuberculosis* sRNA\_1096 and sRNA\_1414, and human hsa-mir-21-5p are probably the links to explain why the pulmonary tuberculosis is a risk factor in developing lung cancer. Among the 45 human miRNAs that are deregulated both in both the diseases (Supplementary Table S1, see supplementary data) and 120 reported *M. tuberculosis* sRNAs in the *M. tuberculosis* genome, we found that there are sequence similarities among human hsa-mir-21-5p and *M. tuberculosis* sRNA\_1096 and sRNA\_1414 (Supplementary Table S2, see supplementary data).

The oncomiR hsa-mir-21 is frequently upregulated in lung cancer [47-55] while it is downregulated in CD4<sup>+</sup> T cells in tuberculosis patients [56]. However, hsa-mir-21 is found upregulated in the host during *M. bovis* BCG infection [57] (Supplementary Table S1, see supplementary data). It is also observed that mir-21 plays a role in T-cell immunity against *M. tuberculosis* [56] and found to affect the anti-mycobacterial T-cell response through targeting IL12 and BCL2 [57]. Reports suggest that the bacteria induced carcinogenesis occurred in multiple ways. These mechanisms include induction or interference of chronic inflammatory and other signalling cascades including TLRs (Toll-like receptors) signalling and acetaldehyde metabolism pathways in various cancers [58-60]. TLRs signalling are generally involved in innate and adaptive immune responses. However, activation of TLRs signalling promotes tumor cell proliferation, growth, invasion, and metastasis [61]. In lung cancer, activation of TLR7 and TLR8 increases survival and chemoresistance of the tumor cells; therefore these two TLRs could be targets for tumor immunotherapy [62]. In *Mycobacterial* infection, TLRs signalling regulates host innate and inflammatory responses and determines the disease outcome [63]. Polymorphisms and over expression of TLR8 is associated with pulmonary tuberculosis susceptibility and infection, respectively [64]. However, the precise mechanism of TLR8 in pulmonary tuberculosis is not yet

known [65]. Fabbri *et al* in 2012 first reported the novel mechanism of oncomiR mir-21 that can act as a ligand for TLR8 to induce inflammatory response leading to tumor growth and metastasis [66]. This study also showed that the “GUUG” motif miR-21 directly binds to TLR8 to induce the TLR-mediated prometastatic inflammatory response.

#### *M. tuberculosis* sRNA\_1096

In our analysis, we found that the GTTG/ GUUG sequence is conserved in hsa-miR-21 and experimentally validated *M. tuberculosis* sRNA\_1096 (**Supplementary Table S2, see supplementary data**). Therefore, it may be implicated that the *M. tuberculosis* sRNA\_1096 may bind to TLR8 in a similar way as miR-21 does through its GUUG motif and thus may activate immune and inflammatory responses and explain a role of TLR8 in pulmonary tuberculosis.

Further, as per our analysis, *M. tuberculosis* sRNA\_1096 may play a role in two-component system pathway and may regulate PE PGRS family and membrane located proteins. PE PGRS family proteins act as variable surface antigens and are involved in multiple levels of the infectious process, modulation of innate immune responses [67], and virulence [68] in *M. tuberculosis*. Similarly, the two-component system is crucial in *Mycobacterial* survival and pathogenicity [69]. In *M. tuberculosis* genome, “CCGTCACCGTTG” sequence of the sRNA\_1096 is present in Arabino-syl transferase EmbC (embC) gene, which is involved in bacteria-host interactions and also modulates immune response in *M. tuberculosis* [70]. Therefore, *M. tuberculosis* sRNA\_1096 could play an important role in *M. tuberculosis* pathogenesis leading to pulmonary tuberculosis. In support to our “Genetic remittance” hypothesis, we predicted that if the *M. tuberculosis* sRNA\_1096 is present or predisposes to human, it may modulate human SH3GL1 (SH3-domain GRB2-like 1) which plays a role in endocytosis [71], regulation of cell cycle in leukaemia [72], positive regulation of cell proliferation and inhibition of apoptosis in multiple myeloma [73], and oncogenesis in gliomas [74]. Hence, sRNA\_1096 may play a critical role in lung cancer risk. Further, if the sRNA\_1096 acts as a ligand to TLR8 similar to mir-21, upon remittance to the host, it may activate TLR8 pathway and along with increasing risk it may also regulate chemoresistance [62] in lung cancer individually or in combination with mir-21. Therefore, we postulate that, the *M. tuberculosis* sRNA\_1096 is involved in pulmonary tuberculosis pathogenesis through multiple infectious processes including TLR8 mediated pathway. The sRNA\_1096 may be transported to host and predisposed during *M. tuberculosis* infection and later acts as ligand to TLR8 through its GTTG/ GUUG sequence similar to mir-21 to activate TLR8 mediated prometastatic pathways and chemoresistance in lung cancer. Similar to

SH3GL1, it may also regulate tumorigenic inflammatory response and cell cycle, respectively leading to lung cancer. Hence, sRNA\_1096 may be an emerging marker for tuberculosis and lung cancer risk and chemoresistance in combination with mir-21.

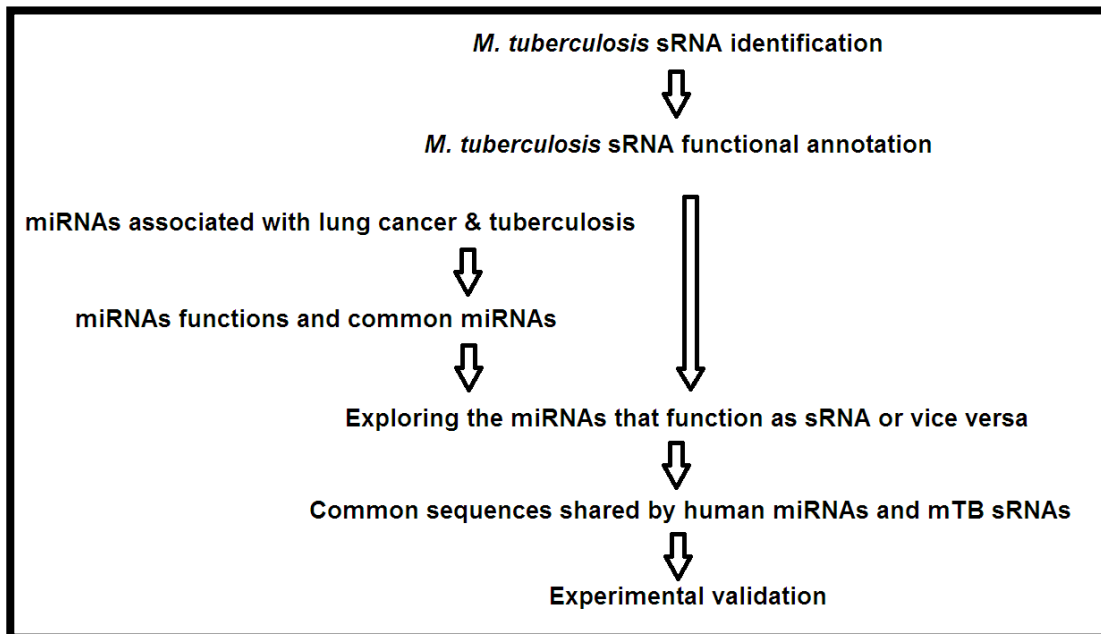
#### *M. tuberculosis* sRNA\_1414

On the other hand, we observed ATCAG/ AUCAG as the common sequence between hsa-mir-21-5p and *M. tuberculosis* sRNA\_1414 (**Supplementary Table S2, see supplementary data**). The sRNA\_1414 is predicted to target glycyl-tRNA synthetase (glyS), which is an essential gene in *M. tuberculosis*. Further, we observed that sRNA\_1414 might regulate drug transporter activity. Aspartyl-tRNA synthetase and tyrosyl-tRNA synthetase are important drug targets in *M. tuberculosis* [75, 76] and polymorphisms in aspartyl-tRNA synthetase is associated with drug resistance mechanism in this pathogen [77]. Although, we predicted glyS is as an essential gene in *M. tuberculosis*, being a human homolog, it is not a suitable target. Thus sRNA\_1414 may probably be involved in regulating the survivability and drug response of the pathogen.

We found sequence matches with human EPS8L1 (EPS8-like 1) and SORBS1 (Sorbin and SH3 domain containing 1) by BLASTn of sRNA\_1414 against human genome. This indicates genetic remittance in tuberculosis associated lung cancer. EPS8L1 encodes a protein that is related to epidermal growth factor receptor pathway substrate 8 and is involved in regulation of Rho protein signal transduction [78], which is associated with small cell lung cancer migration [79]. Similarly, SORBS1 (Sorbin and SH3 domain containing 1) plays an important role in cell-matrix adhesion [80], a key process in cell migration. Therefore, if the *M. tuberculosis* sRNA\_1414 is transferred to the host during the infection, it may lead to lung cancer metastasis in later stage functioning similar to EPS8L1 and SORBS1.

#### Common functions of *M. tuberculosis* sRNA\_1096 and sRNA\_1414

Since the hsa-mir-21 sequence GTTG /GUUA is shared by sRNA\_1096 and ATCAG/ AUCAG by sRNA\_1414 (**Supplementary Table S2, see supplementary data**), we tried to predict the common role of these non-coding RNAs. Our hsa-mir-21 BLASTn against *M. tuberculosis* shows that these GTTG /GUUA and ATCAG/ AUCAG sequences are present in *M. tuberculosis* rpoB gene that provides rifampin/ rifampicin resistance in *M. tuberculosis* [81-84]. Therefore, we presume that all these non-coding RNAs: hsa-mir-21, sRNA\_1096, and sRNA\_1414 could be involved in rifampicin resistance and an up regulation of mir-21 in tuberculosis patient may be a marker of rifampicin resistance.



**Figure 1:** The integrated bioinformatics and experimental validation strategy to elucidate the relationship between tuberculosis and lung cancer linked through non-coding RNAs.

#### Conclusion:

*M. tuberculosis* sRNA<sub>1096</sub> involvement in tuberculosis through multiple molecular processes is of interest to know. This is through the potential activation of TLR8 mediated pro-metastatic inflammatory pathway by acting as a ligand to TLR8. This is similar to mir-21 action leading to lung tumorigenesis and subsequent chemo-resistance. The role of sRNA in cell cycle regulation similar to the human SH3GL1 is relevant. The role of sRNA<sub>1414</sub> in survivability and drug response of the pathogen is contextual. The three non-coding RNAs are predicted to act in rifampicin resistance against *Mycobacterium*. Further data analysis as outlined in Figure 1 including domain/ motif analysis along with experimental validations are required to validate the observation.

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#### Author contributions:

Conceived, designed the experiment, collected and analyzed initial data, coordinated the entire project: DB, Performed all analysis: DB, ST, PG, RK, Wrote the paper: DB, VA guided the project. All authors read and approved the manuscript.

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## Supplementary Data:

### Supplementary Table S1:

Common miRNAs that are deregulated both in pulmonary tuberculosis and lung cancer.

miRNA Name	Regulation in lung cancer	Reference (PMID)/doi	Regulation in Tuberculosis	References (PMID)/doi
hsa-let-7g	Down-regulated	doi: 10.4046/trd.2009.67.5.413, 16461460; 18766170	Up-regulated (serum)	21998423
hsa-let-7i	Down-regulated	18766170	Up-regulated	21998423
hsa-miR-21	Up-regulated	19228723, 19273703, 18766170, 18719201, 21116241, 19493678, 16530703, 16461460, 21351266	Down-regulated Up-regulated in Mycobacterium bovis BCG infection	23613882, 22710123
hsa-miR-23b	Down-regulated	2907339	Up-regulated	21998423
hsa-miR-23b	Up-regulated	18766170	Deregulated	
hsa-miR-22	Down-regulated	2907339; 18766170	Up-regulated	21998423
hsa-miR-26a	Up-regulated	18766170	Down-regulated	23613882
hsa-miR-26a			Up-regulated	22087245
hsa-miR-26b	Down-regulated	18766170	Deregulated	doi:10.4236/jtr.2013.12005
hsa-miR-29a	Down-regulated	2907339; 17890317	Down-regulated	23613882
hsa-miR-29a			Up-regulated	23613882; 21998423; 21785411
hsa-miR-30c	Down-regulated	doi: 10.4046/trd.2009.67.5.413	Up-regulated	21998423
hsa-miR-99b	Up-regulated	16461460; doi: 10.4046/trd.2009.67.5.413	Up-regulated	23233675
hsa-miR-101	Up-regulated	2907339	Up-regulated	21998423
hsa-miR-101	Down-regulated	18766170, doi: 10.4046/trd.2009.67.5.413		
hsa-miR-101	Down-regulated	16530703		
hsa-miR-103	Up-regulated	2907339; 18766170	Up-regulated	21998423
hsa-miR-125a-5p	Down-regulated	20569443	Up-regulated	23233675
hsa-miR-125a-5p	Down-regulated	20569443, 18766170		
hsa-miR-125a-5p	Down-regulated	16530703; 19584273, doi: 10.4046/trd.2009.67.5.413		
hsa-miR-125b	Up-regulated	18766170, doi: 10.4046/trd.2009.67.5.413	Up-regulated	23448104; 21998423; 17911593
hsa-miR-133a	Down-regulated	doi: 10.4046/trd.2009.67.5.413	Up-regulated	22003408
hsa-miR-134	Up-regulated	18766170	Up-regulated	23272999; 22087245

hsa-miR-142-3p	Up-regulated	2907339	Down-regulated	23613882
hsa-miR-142-3p	Down-regulated	19228723, doi: 10.4046/trd.2009.67.5.413		
hsa-miR-143	Down-regulated	19895320, 16530703	Up-regulated	21998423
hsa-miR-144	Down-regulated	doi: 10.4046/trd.2009.67.5.413	Up-regulated	22003408
hsa-miR-146a	Up-regulated	18766170	Up-regulated	21998423
hsa-miR-148b	Up-regulated	2907339	Down-regulated	24084739
hsa-miR-155	Down-regulated	19895320	Up-regulated	21367459; 22712528; 24130493
hsa-miR-155	Up-regulated	16530703, 16461460	Down-regulated	22003408; 21969554
hsa-miR-181b	Up-regulated	18766170	Down-regulated	21998423; 22003408
hsa-miR-191	Up-regulated	18766170, 16461460, 16530703	Up-regulated	21998423
hsa-miR-197	Up-regulated	18766170; 16530703	Up-regulated	24084739
hsa-miR-206	Up-regulated	18766170	Down-regulated	21998423
hsa-miR-210	Up-regulated	20526284, 21116241, 19493678, doi: 10.4046/trd.2009.67.5.413, 16461460, 16530703	Up-regulated	23272999; 22087245
hsa-miR-212	Up-regulated	16530703	Deregulated	doi:10.4236/jtr.2013.12 005
hsa-miR-218	Down-regulated	20838434	Up-regulated	21998423
hsa-miR-222	Down-regulated	2907339, 19895320	Up-regulated	23233675
hsa-miR-222	Up-regulated	19962668, 18766170		
hsa-miR-223	Down-regulated	19895320	Up-regulated	24084739; 22003408
hsa-miR-223	Up-regulated	18766170		
hsa-miR-296-5p	Down-regulated	doi: 10.4046/trd.2009.67.5.413	Down-regulated	24084739
hsa-miR-371-3p	Up-regulated	doi: 10.4046/trd.2009.67.5.413	Down-regulated	21998423
hsa-miR-375	Up-regulated	18766170, 21351266	Up-regulated	21998423
hsa-miR-382	Up-regulated	18766170	Up-regulated	21998423
hsa-miR-423-5p	Upregulated	18766170	Up-regulated	22087245
hsa-miR-432	upregulated	18766170	Up-regulated	23272999; 22087245
hsa-miR-433	Up-regulated	18766170	Up-regulated	21998423
hsa-miR-451	Down-regulated	18766170	Down-regulated	24084739; 22003408
hsa-miR-483-5p	Up-regulated	18766170	Up-regulated	21998423
hsa-miR-486-5p	Down-regulated	21116241, 20194856	Up-regulated	22003408
hsa-miR-501-3p	Up-regulated	18766170	Down-regulated	24084739



hsa-miR-574-5p	Up-regulated	21258252	Up-regulated	21998423
hsa-miR-629	Up-regulated	18766170	Up-regulated	24084739
hsa-miR-744	Up-regulated	18766170	Down-regulated	24084739
hsa-miR-744			Up-regulated	21998423

**Supplementary Table S2:**

Sequences and similarities among of *M. tuberculosis* sRNA\_1096, sRNA\_1414, and human hsa-miR-21

**M. tuberculosis sRNA\_1096**

CGAGCCGTCACC**GTTG**TGCATCGAAAGAGGTCTGATC

**M. tuberculosis sRNA\_1414**

GGCAGACGCGCGCAGCCCGACACGACTACGCGCAAAC**ATCAG**TCA

**hsa-miR-21-5p**

TAGCTT**ATCAG**ACTGAT**GTTG**A

The GUUG / GTTG sequence of mir-21 is involved in binding with TLR8 (PMID: 22753494)

**hsa-miR-21-5p**

UAGCUUAUCAGACUGAU**GUUG**A

**Supplementary Table S3: (Available with author)**

RNApredator based predicted targets of *M. tuberculosis* sRNA\_1096 and sRNA\_1414.